

the organ-on-a-chip company

Human Tissue Models for Better Therapies

Dorota Kurek

PoLiMeR lecture

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E Lecture overview

- In vitro tissue culture
- Limitations of current in vitro models
- Introduction to the 3D organ-on-a-chip
- Building 3D tissues:
 - Modeling gut in vitro
 - Adding complexity

E A leap forward in physiological relevance

The challenge of reductionism:

"Make things as simple as possible, but not simpler", Albert Einstein



Cell culture last decades



3D culture techniques

- a) Scaffold Based
 - Polymeric Hard Scaffolds
 - Biologic Scaffolds
 - Micropatterned Surface Microplates
- b) Non Scaffold Based
 - Hanging Drop Microplates
 - Spheroid Microplates containing Ultra-Low Attachment (ULA) coating
 - Microfluidic 3D cell culture

c) Bioreactors



A BORNER

Non Scaffold Based



3D Cell Culture Gels









3D Microtissues – Terminology and Definitions



Spheroid: HepG2

Spheroid

A spheroid is a 3D cellular aggregate composed of one or more cell types that grow and proliferate, and may exhibit enhanced physiological responses but do not undergo differentiation or self-organization. They are typically derived from primary tissues or immortalized cell lines.



Organoid

"An 'organoid' is a 3D structure derived from either pluripotent stem cells (PSCs), neonatal tissue stem cells, or adult stem and progenitor cells, in which cells spontaneously self-organize into properly differentiated functional cell types and progenitors, and which resemble their *in vivo* counterpart and recapitulate at least some function of the organ."

PSC-derived organoid

Huch M, Koo BY (2015) Modeling mouse and human development using organoid cultures. Development.

3D cell culture



Why do we need better 3D models?

Maximizing the Drug Discovery Pipeline With 3D Models



- Bring biological relevance upstream in the process
- Improve in vitro predictivity/problemsolving capability

- Use 3D cultures to confirm 2D culture results
- Recapitulate human physiology more closely
- Reduce number of in vivo investigative studies

Crgan-on-a-Chip



Dan Huh et al. Science 2010

Multi-organ Chip







www.tissuse.com

Towards a Human-on-a-Chip?









What are the limitations of the current models?



What are the limitations of the current models?



What are the limitations of the current models?



Model development: relevant parameters

- Culture vessel type
- Readout assays
- Tissue selection
- Tissue complexity
- Cell source
- Extracellular matrix
- Seeding density

- Medium composition
- Seeding orientation
- Flow settings

Microfluidics in OrganoPlate®



 3-Dimensional culture

- Flow
- Multiple cell types
- Fully compatible

■ PhaseGuide[™] Technology

PhaseGuide™



Vulto et al 2011 Phaseguides: a paradigm shift in microfluidic priming and emptying. *Lab Chip* 11(9) 1596-1602



https://www.youtube.com/watch?v=BhFETKQqJY0

iPS-derived 3D neuronal networks





Rapid network formation directly after seeding in OrganoPlate®



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Wevers, N. R. et al. Sci. Rep. 6, 38856 (2016).

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iPSC-derived 3D neuronal networks



Inducing seizure





Synchronicity observed in 4-AP exposed cultures (day 19)



How to set up 3D organotypic culture models?

Model development - In vitro Gut



- The gut is part of the gastrointestinal tract
 - Large intestine (cecum, colon, rectum, and anus)
 - Small intestine (duodenum, ileum, and jejunum)
 - Important for absorption of orally delivered drugs
- Dysfunction involved in many different diseases
 - Inflammatory Bowel Disease (IBD)
 - Acute and chronic enteritis/ colitis
 - Peptic ulcers (affecting the duodenum)
 - Coeliac disease
 - Whipple's disease
 - Cancer
 - &&&

Model development: required organ complexity

"Make things as simple as possible, but not simpler", Albert Einstein

- Depends on your research question
 - E.g. Colon barrier integrity upon toxicant stimuli
 - Or Disease modelling: Inflammatory Bowel Disease (IBD)



Continuous passive perfusion (bidirectional)

 OrganoPlates[®] are placed on top of the rocker platform inside the incubator





Growing Tubules in the OrganoPlate [®] 3-lane





Growing Caco-2 gut tubules

 Caco-2 intestinal model forms tubular structures with accessible apical and basal sides, expressing various drugs transporters









Gut - 2D vs 3D gene expression

 Caco-2 cells cultured in the OrganoPlate[®] show upregulation of a myriad of gut markers compared to Caco-2 cells cultured in 2D



Caco-2 gut tubes in OrganoPlates®



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Staurosporine-induced gut toxicity





CrganoTEER – barrier assessment



Disease modelling – gut inflammation

Inflammatory Bowel Disease (IBD)



Disease modelling – gut inflammation

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T+

D4-D11

Beaurivage et al 2019 Int J Mol Sci Galápagos

Control gut tubules

+ inflammatory trigger (cytokines)

Control gut tubules

+ inflammatory trigger 72h



Inflamed Gut-on-a-Chip Model for Drug Discovery



- Adenoviral Knockdown of Inflammatory Effectors Prevents IBD-like Phenotype in Caco-2
- Exposure to anti-inflammatory TPCA-1 Prevent the Inflammatory State of Caco-2 Tubules
 PoLiMeR lecture, 2020

An intestine-on-a-chip model of plug and play modularity to study inflammatory processes



Gijzen et al 2020 SLAS Tech PoLiMeR lecture, 2020

Adding more complexity to gut model

Disease characteristic

Multiple cell interactions and readouts

Gut model with intestinal tube, stroma and endothelial vessel containing immune components

Intestinal epithelial and endothelial barrier assessment in OrganoPlate

Intestinal barrier



Gut tubule (enterocytes and goblet cells)

- Intestinal myofibroblasts
- Vasculature with immune cells (macrophages and dendritic cells)





Human Colon Organoid tubules in OrganoPlate



- 3D perfused hC tubule formation with apical and basal access



IBD modelling in colon organoid tubules





Gut tubules IL8 release





Hubrecht Institute

Blood vessel in the OrganoPlate® 3-lane



Day 2 Day 3 Angiogenic sprouting Fusion of capillaries and merging and lumen formation

Lumen widening

Angiogenic Sprouts

Staining legend: DNA VE-Cadherin Actin

Max. projection

Orthogonal view

Z-stack

PoLiMeR lecture, 2020

Van Duinen et al. 2018. Angiogenesis

1284

CrganoPlate [®] Graft







Membrane-free

Pump-free perfusion

PoLiMeR lecture, 2020

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- Complex co-cultures
- Excellent imaging

E Phase 1: 3D tube formation with Endothelial Cells



E Continuous perfusion





Phase 2: Vascular bed formation



E Phase 3: *Tissue transplantation*







Compatible with automation



64 individual microtissues ready for vascularization



Tissue vascularization in OrganoPlate Graft®





with vessels

E Liver microtissue vascularization in OrganoPlate Graft



 Sectioning of liver microtissues recovered from OrganoPlate Graft and 3D imaging show vessels penetrating liver microtissues
 POLIMER lecture, 2020

Z Vessel stabilization in co-culture with microtissues



Reproducibly stable vessels over the subsequent 21 days of co-culture with microtissues

DILI in vascularized liver tissue



- Vascularised liver microtissue is more sensitive to compound A
- Protective function liver microtissue on vascular tox

Connecting patient tissue with human vasculature

- Applications: vascularizing xenografts, organoids, spheroids



Tumor tissues in the OrganoPlate Graft

Glioma spheroids

HNSCC organoids

Ovary Cancer explant





- Glioma OMS interact with vascular bed, cells migrating into the ECM
- Head-and-neck organoids and vascular bed interaction
- Ovary cancer explant shows massive interaction with vascular beds.







Take home message

- Organotypic models recapitulate human physiology
- Improve in vitro predictivity
- Reduce number of in vivo investigative studies



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the organ-on-a-chip company



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